



Sorting by pools: Decreasing pool diversities

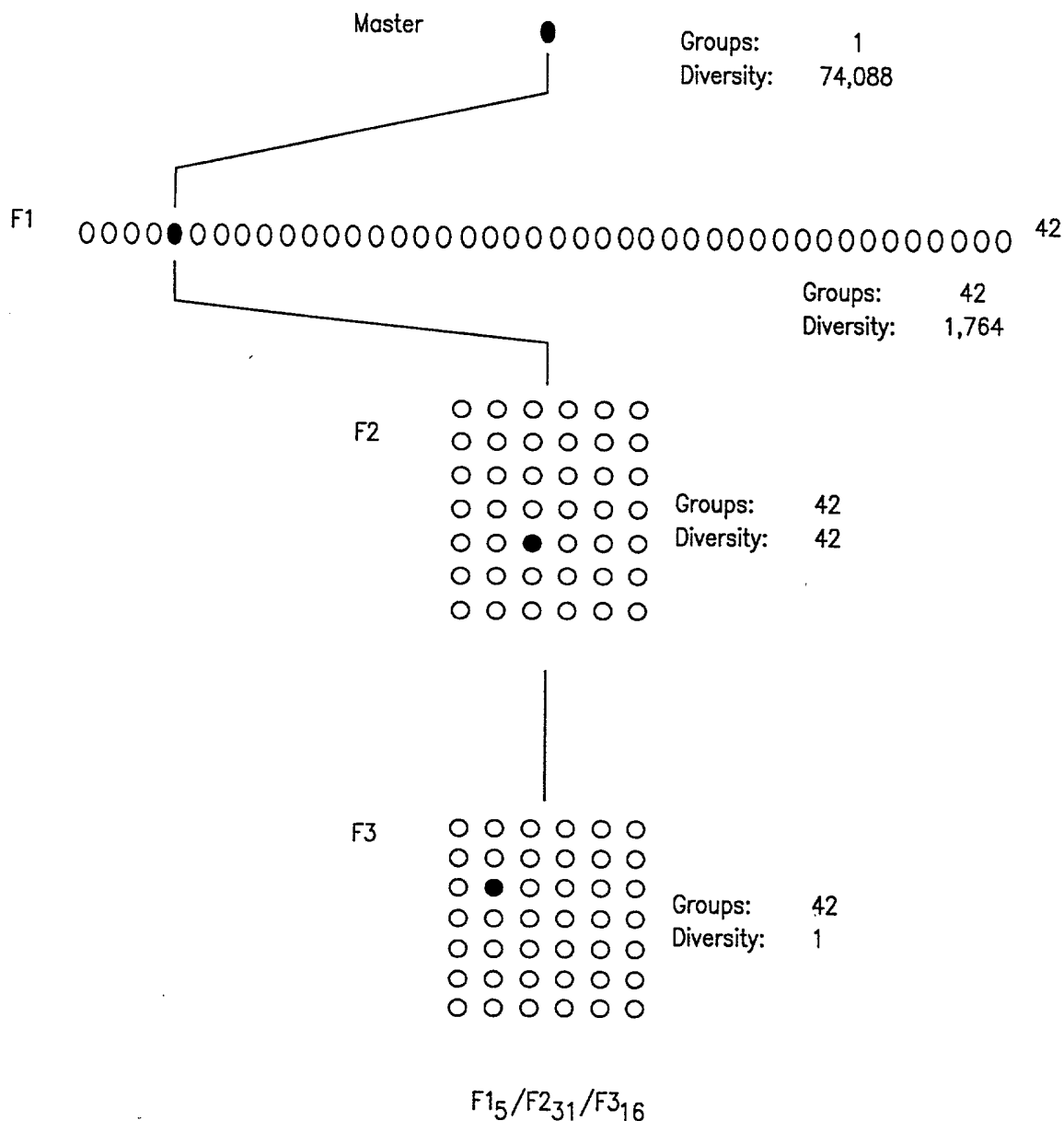


FIG. 2

Sorting by pools: Screening large diversity libraries

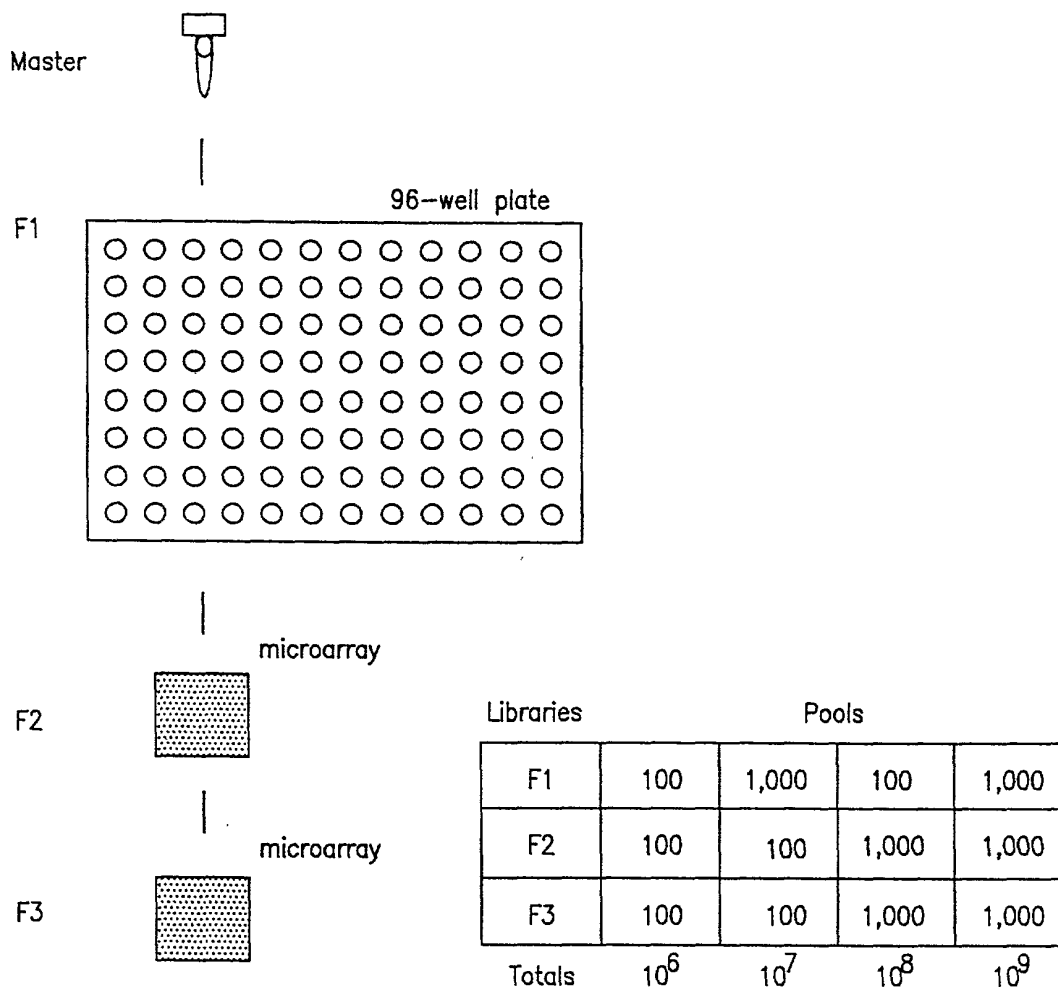


FIG. 3

Searching a mutation library

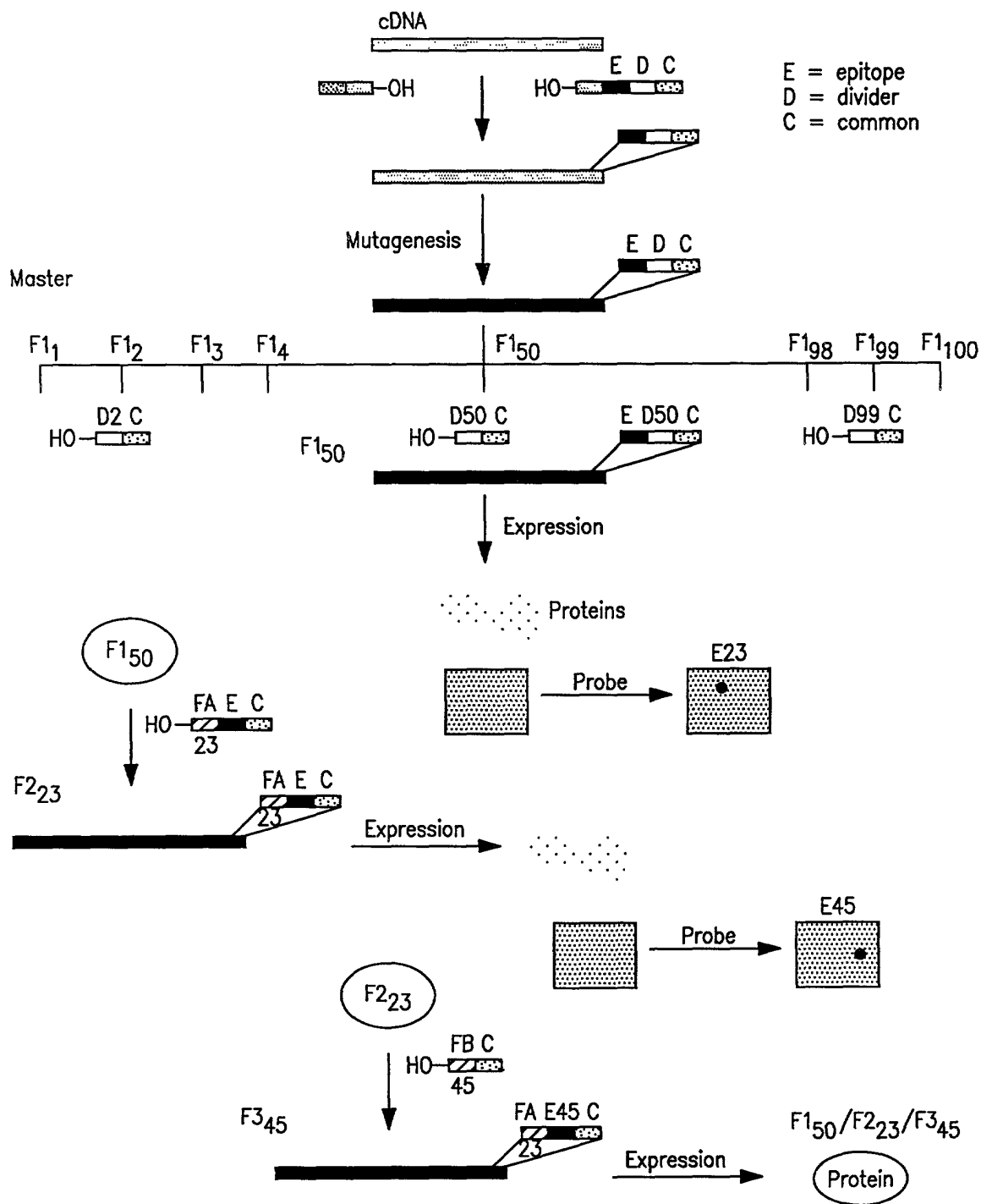
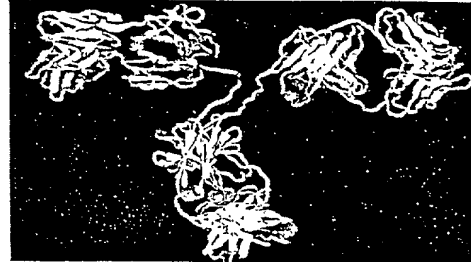
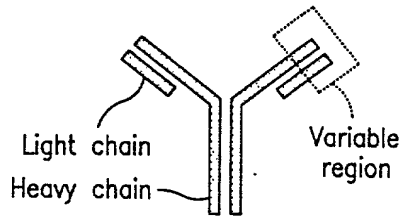
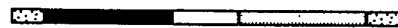
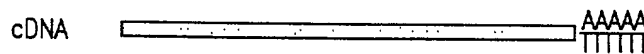


FIG. 4

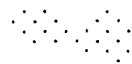
Making a recombinant antibody library



Spleen cells or PBLs



Expression



Antibodies

FIG. 5

Creating the master antibody library: Primer incorporation

1. mRNA purification from spleen or PBLs

2. cDNA synthesis

3. amplification

4. assembly

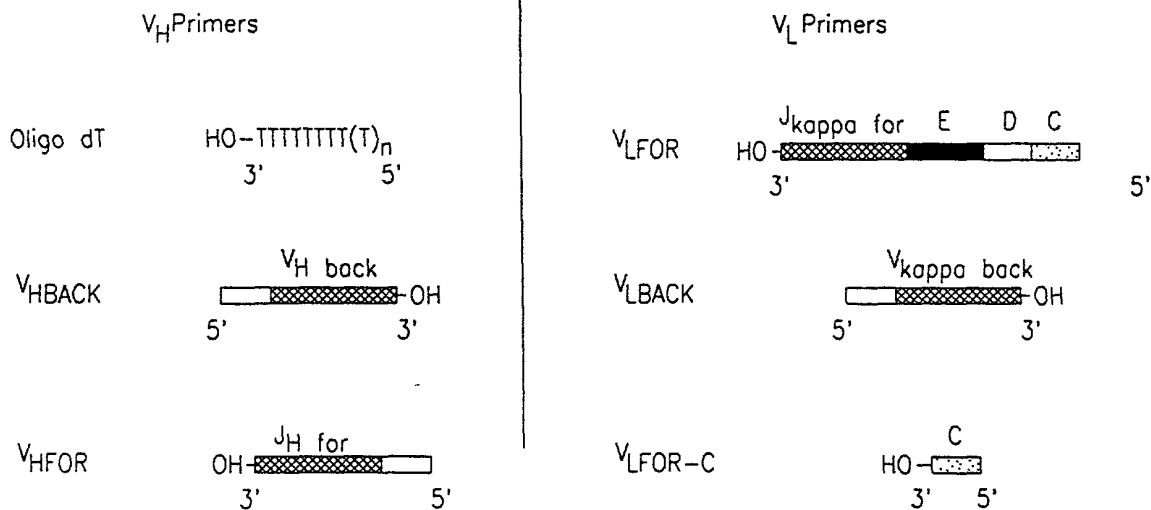
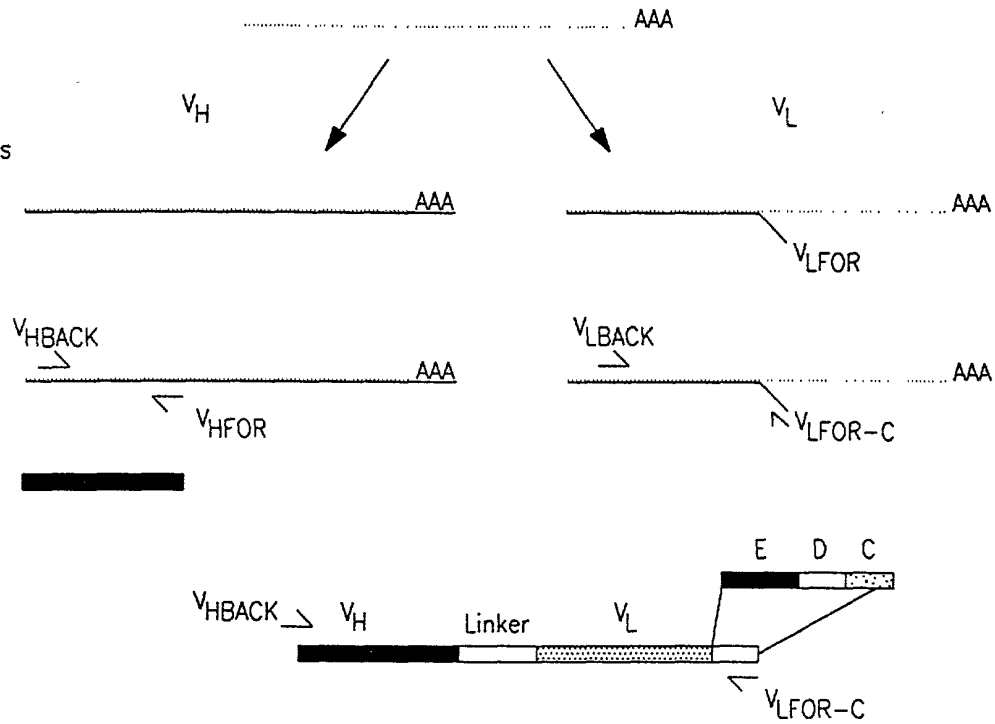


FIG. 6

Creating the master antibody library: Linker addition

1. mRNA purification from spleen or PBLs

2. cDNA synthesis

3. amplification

4. assembly

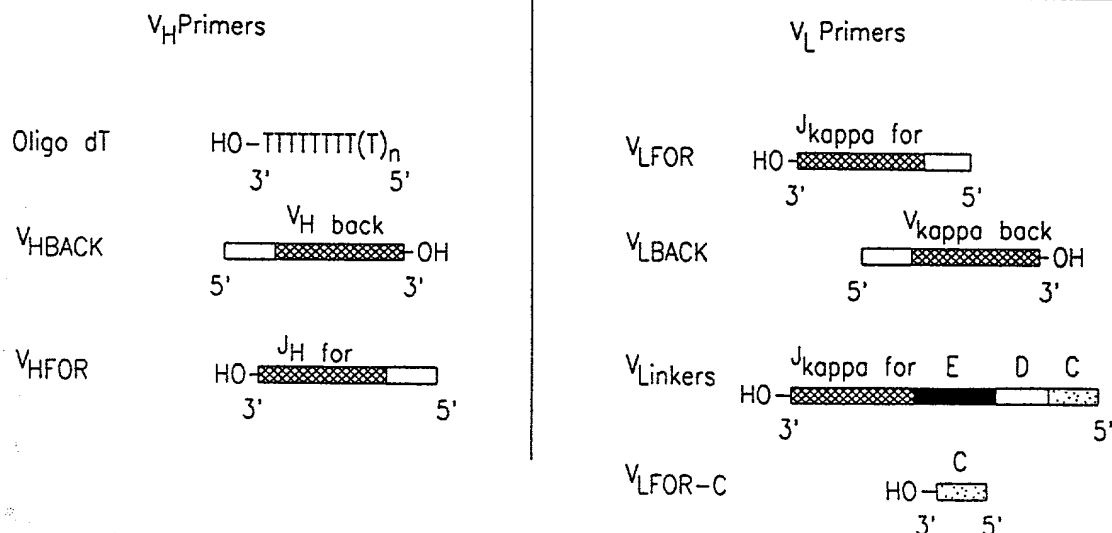
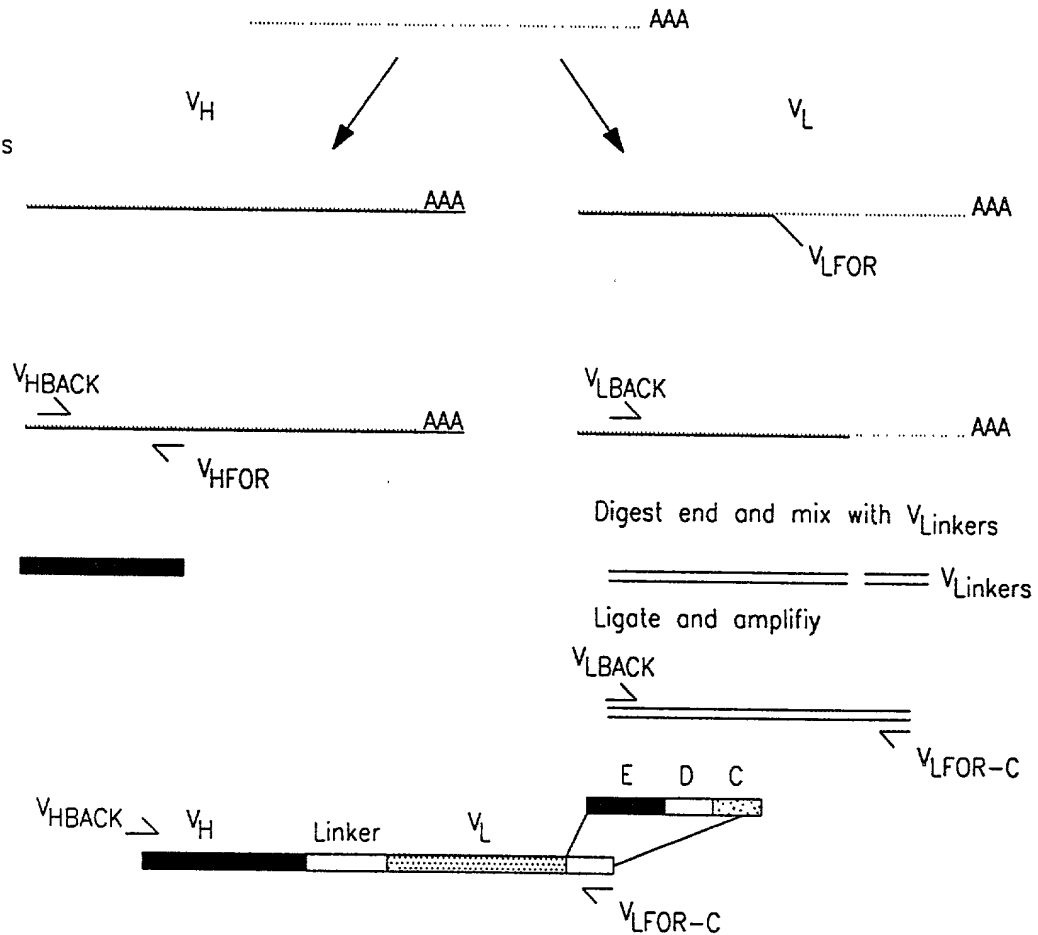
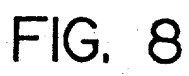


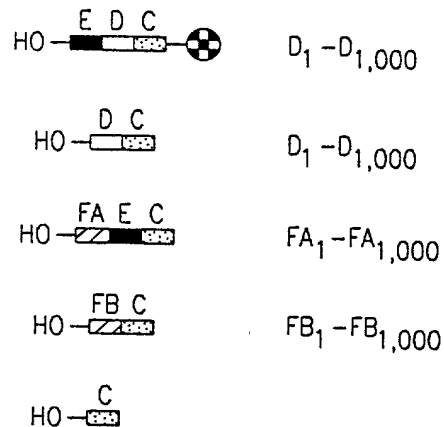
FIG. 7



Physical elements to include in the kits and combinations

- *Anti-tag Arrays*TM

- Primer sets

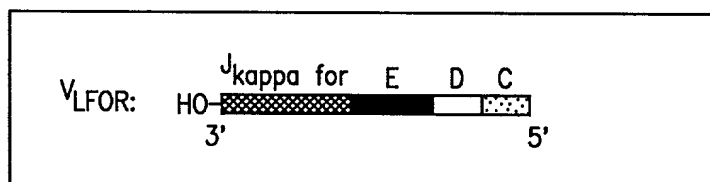


- Readers

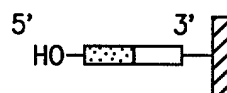
- Software

FIG. 9

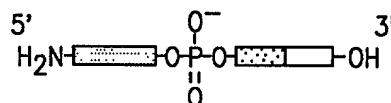
Making the V_LFOR primers: Solid phase synthesis



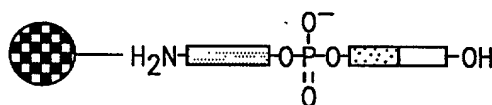
1. Synthesize oligo on solid support



2. Add aminolink prior to cleavage



3. Couple to tosyl activated magnetic beads



4. Extended by hybridizing with DNA patch and ligating

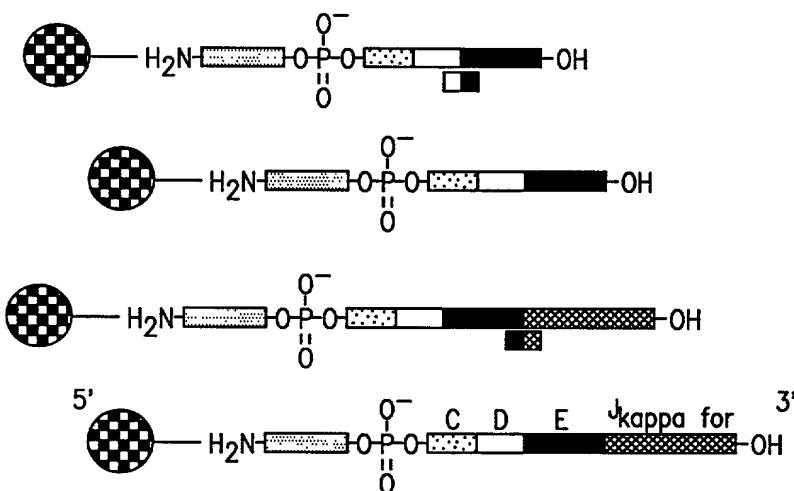
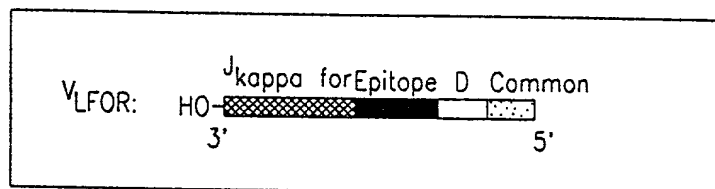
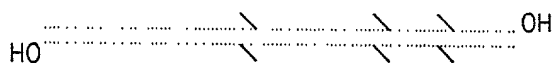


FIG. 10

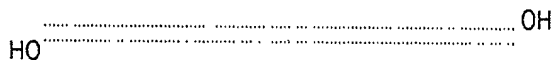
Making the V_LFOR primers: Overlapping hybridization



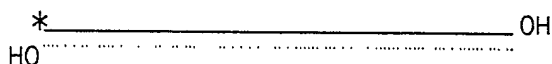
- Synthesize 4,028 different oligos:
 (26 for J_{kappa} for ; 2,000 for Epitope, 2,000 for D; 2 for Common)
- 2. Assemble oligos for + and - strands of the different regions



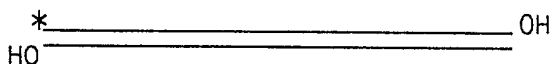
- 3. Ligase the assembled oligos



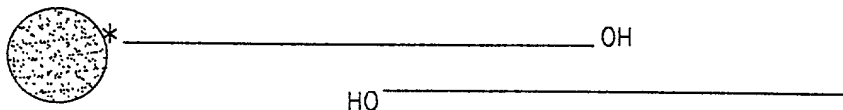
- 4. 1st strand synthesis with biotinylated primer



- 2nd strand synthesis with non-biotinylated primer



- 6. Bind to avidin coated magnetic beads and then denature



- 7. Purify non-biotinylated ssDNA

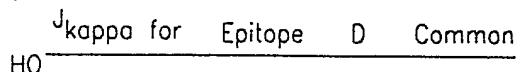


FIG. 11

Building the collection of antibody/tag pairs: Hybridoma screening

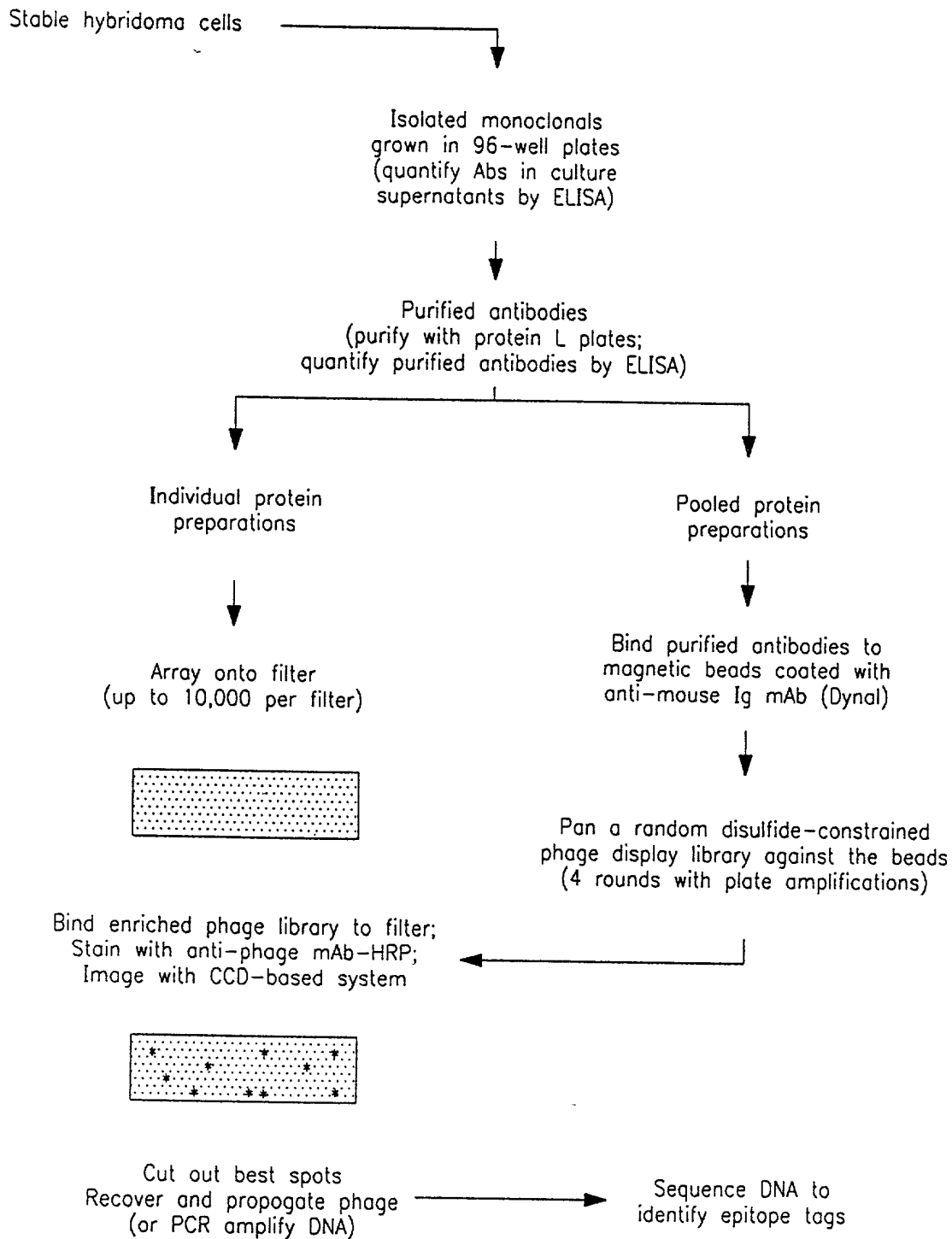


FIG. 12

Table 3 Primers for PCR Amplification of Human Antibody Variable Regions (V genes)

1. V gene primary PCR

A. Human VH back primers (sense)

HuVH1aBACK	5'-CAG GTG CAG CTG GTG CAG TCT GG-3'
HuVH2aBACK	5'-CAG GTC AAC TTA AGG GAG TCT GG-3'
HuVH3aBACK	5'-GAG GTG CAG CTG GTG GAG TCT GG-3'
HuVH4aBACK	5'-CAG GTG CAG CTG CAG GAG TCG GG-3'
HuVH5aBACK	5'-GAG GTG CAG CTG TTG CAG TCT GC-3'
HuVH6aBACK	5'-CAG GTA CAG CTG CAG CAG TCA GG-3'

B. Human JH forward primers (anti-sense)

HuJH1-2FOR	5'-TGA GGA GAC GGT GAC CAG GGT GCC-3'
HuJH3FOR	5'-TGA AGA GAC GGT GAC CAT TGT CCC-3'
HuJH4-5FOR	5'-TGA GGA GAC GGT GAC CAG GGT TCC-3'
HuJH6FOR	5'-TGA GGA GAC GGT GAC CGT GGT CCC-3'

C. Human V kappa back primers (sense)

HuVk1aBACK	5'-GAC ATC CAG ATG ACC CAG TCT CC-3'
HuVk2aBACK	5'-GAT GTT GTG ATG ACT CAG TCT CC-3'
HuVk3aBACK	5'-CAA ATT GTG TTG ACG CAG TCT CC-3'
HuVk4aBACK	5'-GAC ATC GTG ATG ACC CAG TCT CC-3'
HuVk5aBACK	5'-CAA ACG ACA CTC ACG CAG TCT CC-3'
HuVk6aBACK	5'-GAA ATT GTG CTG ACT CAG TCT CC-3'

C. Human V lambda back primers (sense)

HuVλ1BACK	5'-CAG TCT GTG TTG ACG CAG CCG CC-3'
HuVλ2BACK	5'-CAG TCT GCC CTG ACT CAG CCT GC-3'
HuVλ3aBACK	5'-TCC TAT GTG CTG ACT CAG CCA CC-3'
HuVλ3bBACK	5'-TCT TCT GAG CTG ACT CAG GAC CC-3'
HuVλ4BACK	5'-CAC GTT ATA CTG ACT CAA CCG CC-3'
HuVλ5BACK	5'-CAG GCT GTG CTC ACT CAG CCG TC-3'
HuVλ6BACK	5'-AAT TTT ATG CTG ACT CAG CCC CA-3'

D. Human J kappa forward primers (anti-sense)

HuJk1FOR	5'-ACG TTT GAT TTC CAC CTT GGT CCC-3'
HuJk2FOR	5'-ACG TTT GAT CTC CAG CTT GGT CCC-3'
HuJk3FOR	5'-ACG TTT GAT ATC CAC TTT GGT CCC-3'
HuJk4FOR	5'-ACG TTT GAT CTC CAC CTT GGT CCC-3'
HuJk5FOR	5'-ACG TTT AAT CTC CAG TCG TGT CCC-3'

D. Human J. lambda forward primers (anti-sense)

HuJλ1FOR	5'-ACC TAG GAC GGT GAC CTT GGT CCC-3'
HuJλ2-3FOR	5'-ACC TAG GAC GGT CAG CTT GGT CCC-3'
HuJλ4-5FOR	5'-ACC TAA AAC GGT GAG CTG GGT CCC-3'

FIG. 13A

2. Linker fragment PCR

F. Reverse JH for scFv linker (sense)

	————FR4 heavy————— ————linker—————
RHuJH1-2	5'-GC ACC CTG GTC ACC GTC TCC TCA GGT GG-3'
RHuJH3	5'-GG ACA ATG GTC ACC GTC TCT TCA GGT GG-3'
RHuJH4-5	5'-GA ACC CTG GTC ACC GTC TCC TCA GGT GG-3'
RHuJH6	5'-GG ACC ACG GTC ACC GTC TCC TCA GGT GG-3'

F. Reverse Vk for scFv linker (anti-sense)

	————FRI light————— ————linker—————
RHuVk1aBACKFv	5'-GG AGA CTG GGT CAT CTG GAT GTC CGA TCC GCC-3'
RHuVk2aBACKFv	5'-GG AGA CTG AGT CAT CAC AAC ATC CGA TCC GCC-3'
RHuVk3aBACKFv	5'-GG AGA CTG CGT CAA CAC AAT TTC CGA TCC GCC-3'
RHuVk4aBACKFv	5'-GG AGA CTG GGT CAT CAC GAT GTC CGA TCC GCC-3'
RHuVk5aBACKFv	5'-GG AGA CTG CGT GAG TGT CGT TTC CGA TCC GCC-3'
RHuVk6aBACKFv	5'-GG AGA CTG AGT CAG CAC AAT TTC CGA TCC GCC-3'

F. Reverse Vλ for scFv linker (anti-sense)

	————FRI light————— ————linker—————
RHuVλBACK1Fv	5'-GG CGG CTG CGT CAA CAC AGA CTG CGA TCC GCC ACC GCC AGA G-3'
RHuVλBACK2Fv	5'-GC AGG CTG AGT CAG AGC AGA CTG CGA TCC GCC ACC GCC AGA G-3'
RHuVλBACK3aFv	5'-GG TGG CTG AGT CAG CAC ATA GGA CGA TCC GCC ACC GCC AGA G-3'
RHuVλBACK3bFv	5'-GG GTC CTG AGT CAG CTC AGA AGA CGA TCC GCC ACC GCC AGA G-3'
RHuVλBACK4Fv	5'-GG CGG TTG AGT CAG TAT AAC GTG CGA TCC GCC ACC GCC AGA G-3'
RHuVλBACK5Fv	5'-GA CGG CTG AGT CAG CAC AGA CTG CGA TCC GCC ACC GCC AGA G-3'
RHuVλBACK6Fv	5'-TG GGG CTG AGT CAG CAT AAA ATT CGA TCC GCC ACC GCC AGA G-3'

3. Pull-through primers for introduction of restriction sites*

G. Human VH back (Sfi)primers (sense)

HuVH1aBACKSf1	5'-GTC CTC GCA ACT <u>GCG GCC</u> CAG <u>CCG GCC</u> ATG GCC CAG GTG CAG CTG GTG CAG TCT GG-3'	————FRI heavy—————
HuVH2aBACKSf1	5'-GTC CTC GCA ACT <u>GCG GCC</u> CAG <u>CCG GCC</u> ATG GCC CAG GTG AAC TTA AGG GAG TCT GG-3'	
HuVH3aBACKSf1	5'-GTC CTC GCA ACT <u>GCG GCC</u> CAG <u>CCG GCC</u> ATG GCC GAG GTG CAG CTG GTG GAG TCT GG-3'	
HuVH4aBACKSf1	5'-GTC CTC GCA ACT <u>GCG GCC</u> CAG <u>CCG GCC</u> ATG GCC CAG GTG CAG CTG CAG GAG TCG GG-3'	
HuVH5aBACKSf1	5'-GTC CTC GCA ACT <u>GCG GCC</u> CAG <u>CCG GCC</u> ATG GCC CAG GTG CAG CTG TTG CAG TCT GG-3'	
HuVH6aBACKSf1	5'-GTC CTC GCA ACT <u>GCG GCC</u> CAG <u>CCG GCC</u> ATG GCC CAG GTA CAG CTG CAG CAG TCA GG-3'	

H. Human J kappa forward (Not) primers (anti-sense)

HuJk1FORNot	5'-GAG TCA TTC TCG ACT <u>TGC GGC CGC</u> ACG TTT GAT TTC CAC CTT GGT CCC-3'	————FR4 light—————
HuJk2FORNot	5'-GAG TCA TTC TCG ACT <u>TGC GGC CGC</u> ACG TTT GAT CTC CAG CTT GGT CCC-3'	

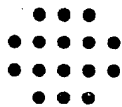
H. Human J kappa forward (Not) primers (anti-sense)(continued)

HuJk3FORNot	5'-GAG TCA TTC TCG ACT <u>TGC GGC CGC</u> ACG TTT GAT ATC CAC TTT GGT CCC-3'	————FR4 light—————
HuJk4FORNot	5'-GAG TCA TTC TCG ACT <u>TGC GGC CGC</u> ACG TTT GAT CTC CAC CTT GGT CCC-3'	
HuJk5FORNot	5'-GAG TCA TTC TCG ACT <u>TGC GGC CGC</u> ACG TTT AAT CTC CAG TCG TGT CCC-3'	

H. Human J lambda forward (Not) primers (anti-sense)

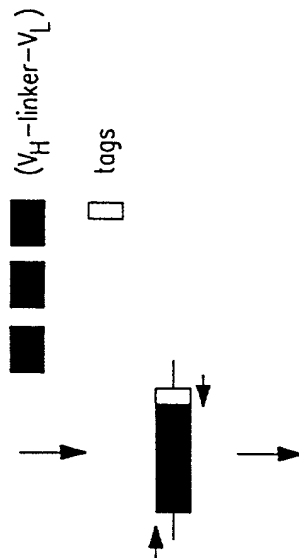
HuJl1FORNOT	5'-GAG TCA TTC TCG ACT <u>TGC GGC CGC</u> ACC TAG GAC GGT GAC CTT GGT CCC-3'	FR4 light
HuJl2-3FORNOT	5'-GAG TCA TTC TCG ACT <u>TGC GGC CGC</u> ACC TAG GAC GGT CAG CTT GGT CCC-3'	
HuJl4-5FORNOT	5'-GAG TCA TTC TCG ACT <u>TGC GGC CGC</u> ACC TAA AAC GGT GAG CTG GGT CCC-3'	

*Recognition site for restriction enzyme is underlined.



step I

Tag and assemble immunoglobulin genes



Create 1,000 sub-libraries by separate PCR amplification reactions using tag-specific PCR primers

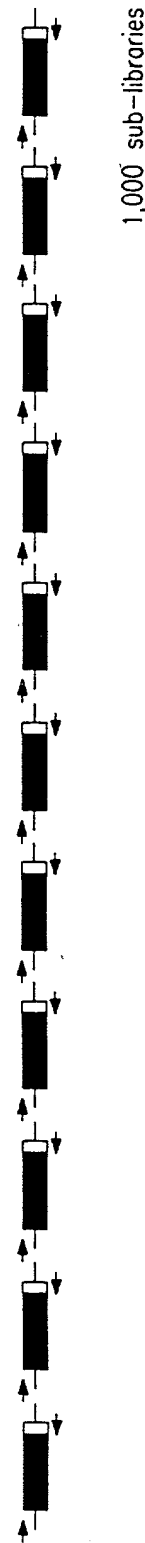
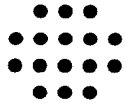


FIG. 14A



step II

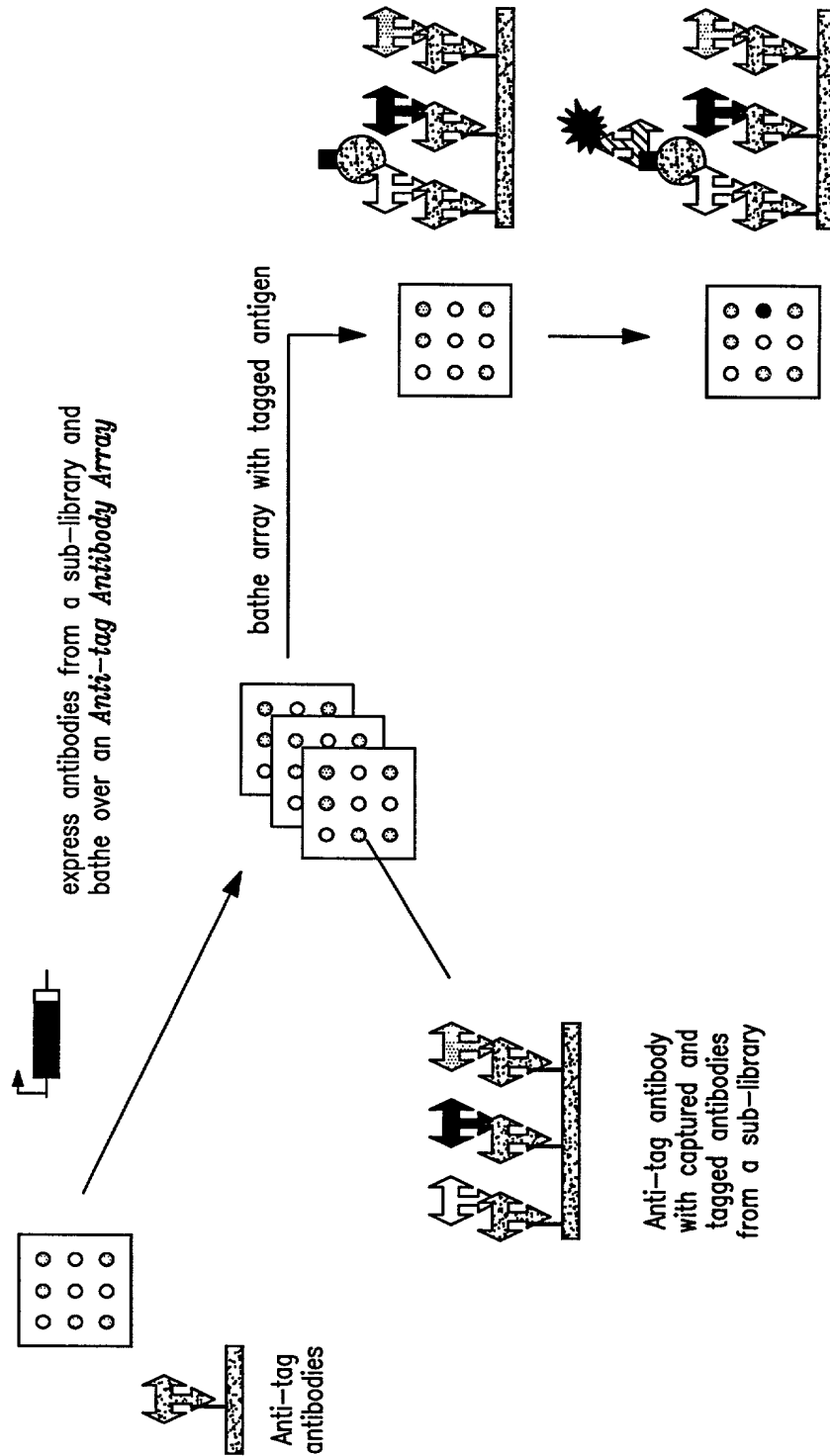
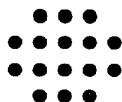


FIG. 14B



step III

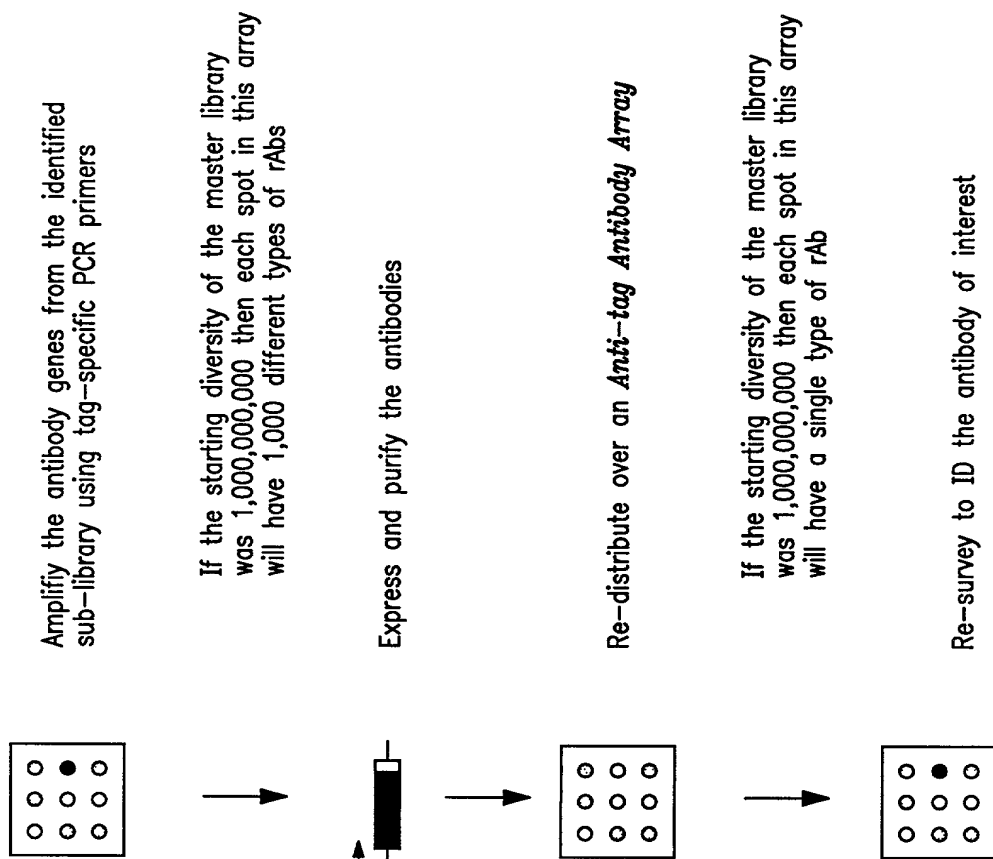
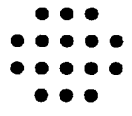
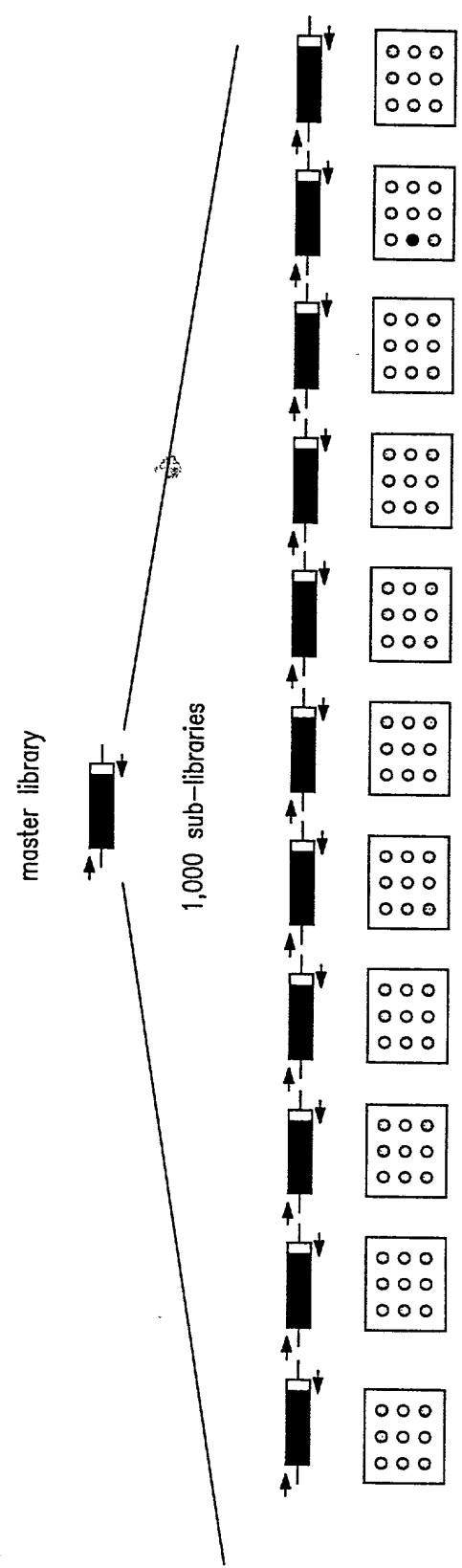


FIG. 14C

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summary



Round	Library diversity	Sub-libraries	Spot diversity	Arrays used
0	1,000,000,000	1,000		
I	1,000,000		1,000	1,000
II	1,000		1	1

FIG. 14D

Modification searches

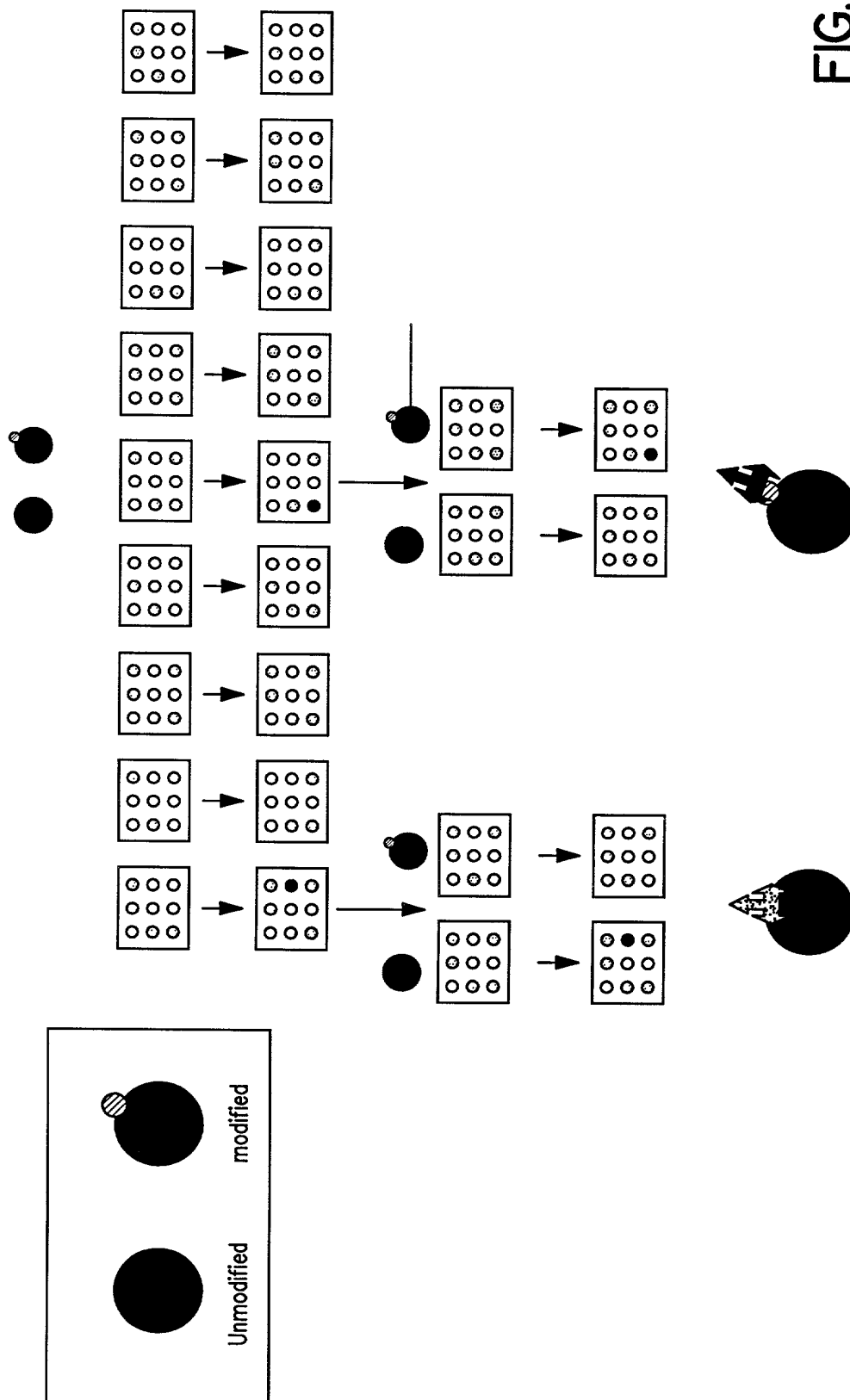
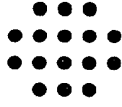
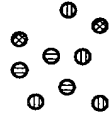


FIG. 15



Simultaneous searches

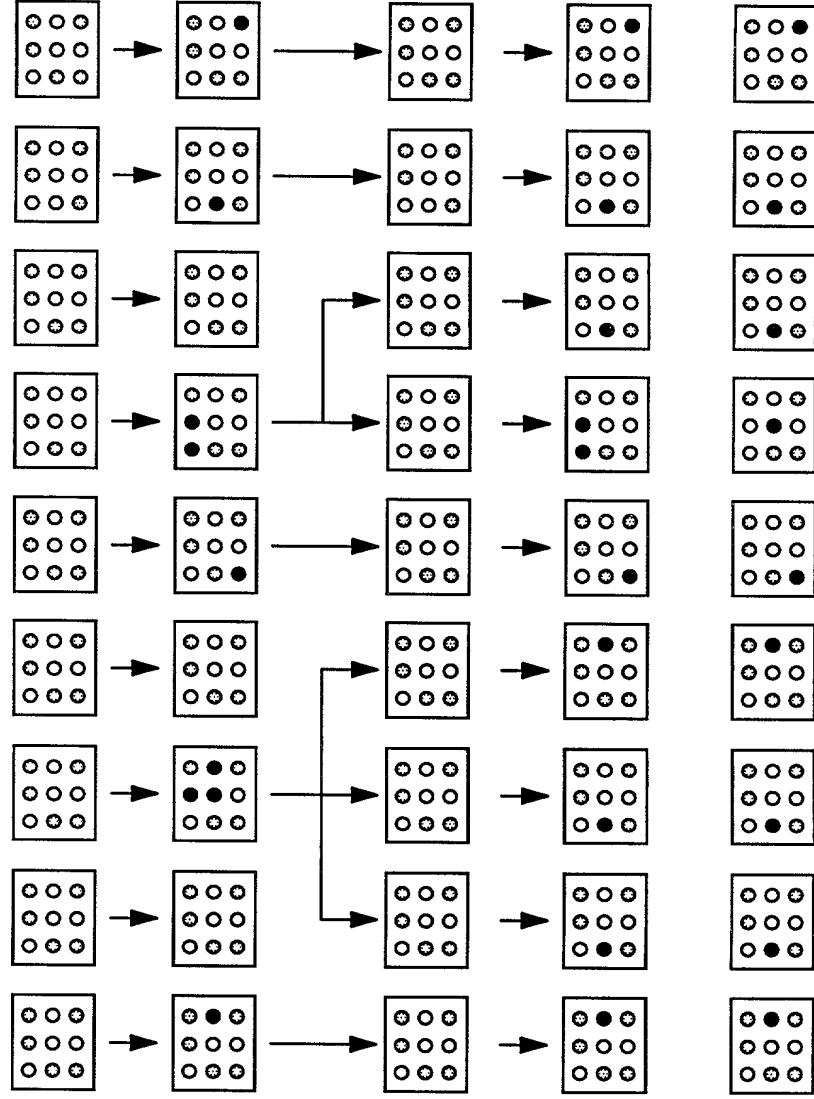


Round Arrays Bait Probe

I 1,000 Abs Ags

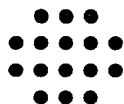
II 1,000 Abs Ags

III 1,000 Abs Abs
3,000

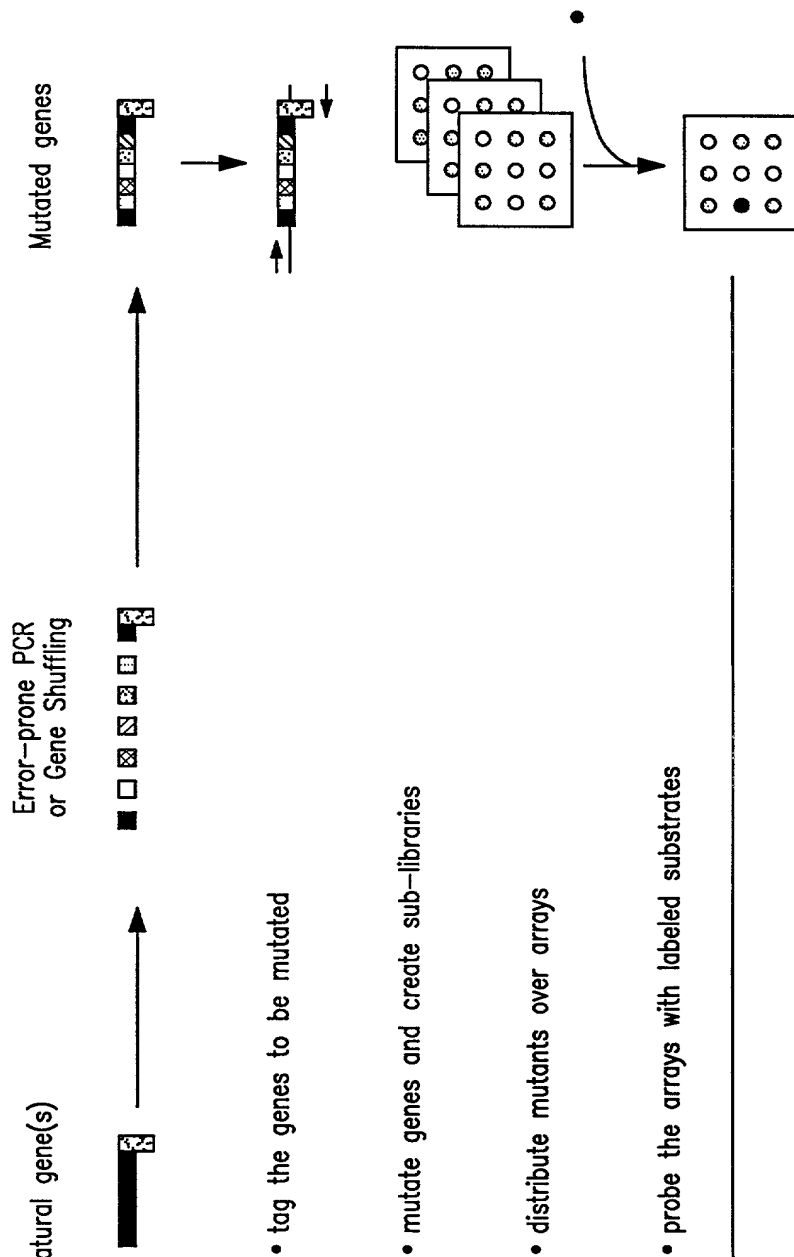


3 Arrays per Ag

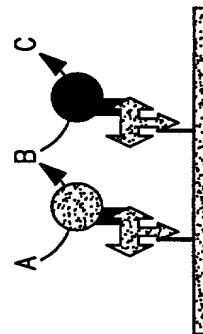
FIG. 16



Protein interaction mapping

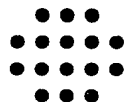


- tag the genes to be mutated
- mutate genes and create sub-libraries
- distribute mutants over arrays
- probe the arrays with labeled substrates



Spots can contain mixtures of enzymes
for detection or pathway engineering

FIG. 17



Protein interaction mapping

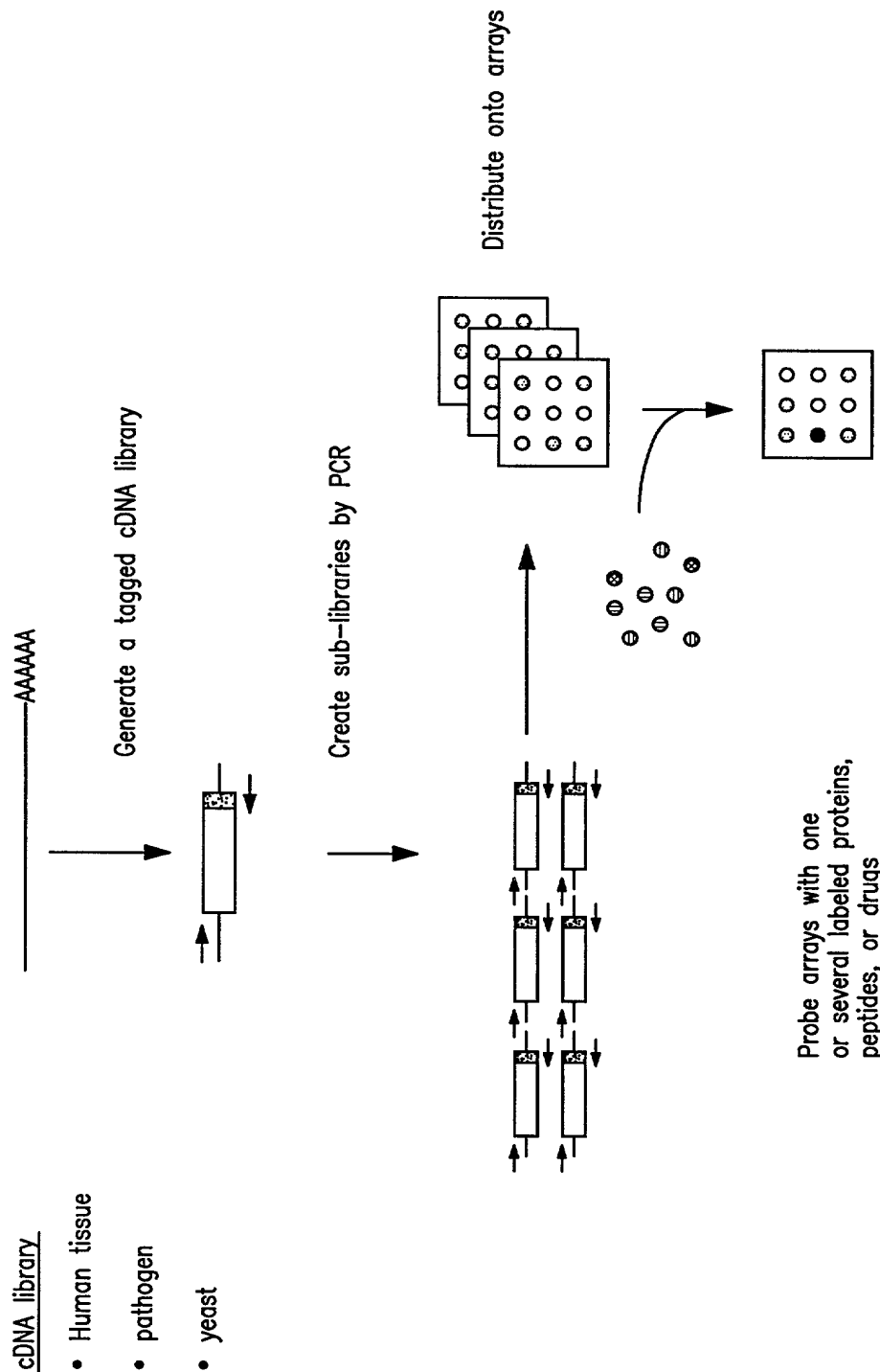


FIG. 18

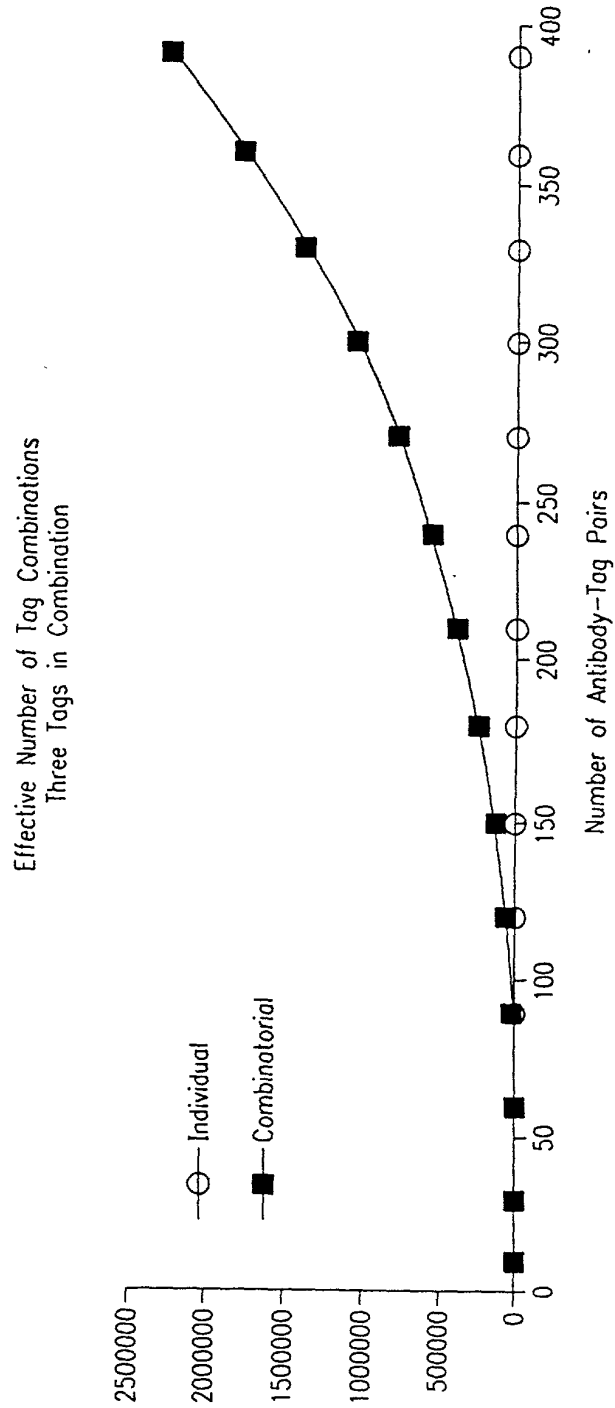


FIG. 19